Genotyping of *Helicobacter pylori* cagA Gene from a Patient Who Failed Eradication Therapy: A Case Report and Review of the Literature

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ABSTRACT

Helicobacter pylori (H. pylori) is a bacterium that causes chronic gastritis, gastric and duodenal ulcers and gastric cancer. Here we report a female patient presenting with dyspepsia. She was tested positive six times by the Rapid Urease test and Urea breath test (UBT). A culture of gastric biopsy was done and the isolate showed resistance to Clarithromycin and Metronidazole while polymerase chain reaction (PCR) revealed the presence of cagA H. pylori virulence gene. Presence of cagA might not be a risk factor in development of Metronidazole resistance to antibiotic therapy. In conclusion, we report a female Malaysian Indian with cagA positive H. pylori infection, but experienced Metronidazole resistance to antibiotic therapy.

KEY WORDS

Helicobacter pylori, CagA, antibiotic resistance

INTRODUCTION

Helicobacter pylori (H. pylori) is a Gram-negative microaerophilic bacterium that causes gastrointestinal diseases such as chronic gastritis, gastric and duodenal ulcers and gastric cancer¹⁻³. H. pylori infection can be diagnosed by invasive (culture, rapid urease test, PCR and histology) and non invasive tests (serology, stool antigen test and ¹³C-urea breath test (UBT)⁴⁻⁶). Invasive test has the advantage of being able to determine antibiotic- susceptibility and H. pylori genotypes⁷⁾.

In order to eradicate *H. pylori* infection, triple therapy using a proton pump inhibitor (PPI) with Clarithromycin and Amoxicillin or Metronidazole is recommended as the first-line treatment regimen. In case the triple therapy fails bismuth-containing quadruple therapy, which involves the inclusion of additional antibiotics to the first-line treatment regimen is used⁸.

In Malaysia, Indians have been found to posses the highest prevalence of infection of about 68.9-75% as compared to Chinese 45.0-60.6% and Malays 8-43.3% 9.10).

A number of *H. pylori* virulence genes, including *cagA*, and *SabA* and have been associated with the most serious clinical outcomes and pathogenic bacteria^{11,12}.

CASE REPORT

This is 41 years old Malaysian female of Indian origin referred from Melaka Hospital for persistent dyspepsia. The patient was positive for *H. pylori* six times by Rapid urease test and UBT. The endoscopic find-

ing was gastritis. She had a history of antiphosphollipid syndrome, bronchial asthma and dystunation uterine bleeding. Despite several treatment regimens the eradication therapy failed. Later on 20/2/2013 the patient underwent an upper gastrointestinal endoscopy at Hospital Kuala Lumpur and two gastric biopsies were taken for culture and sensitivity and Polymerase chain reaction (PCR).

The biopsy sample was put in brucella broth and immediately cultured onto Columbia agar supplemented with 8% sheep blood and dent

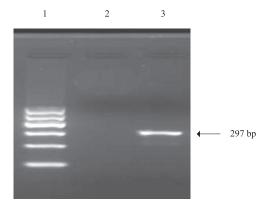


Figure 1. genotyping of cagA gene by PCR: lane 1, 100-bp DNA marker, lane 2 negative control without DNA, lane 3, *H. pylori* cagA positive strain.

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antibiotic. The plate was incubated at 37° C for seven days under microaerophilic conditions. The isolate was confirmed to be *H. pylori* by Gram's stain and positive urease, catalase, and oxidase tests.

Minimum inhibitory concentration was determined by E-test strips. The isolate showed resistance to clarithromycin and metronidazole but was sensitive to Teicoplanin and Levofloxacin.

Genomic DNA extraction

DNA was extracted from biopsy tissue by use of the QIAamp DNA kit (Qiagen, Germany) according to the manufacturer's recommendations and stored at -20°C until analysis.

PCR conditions and amplification of cagA gene

PCR amplification of *cagA* was carried out using two primer sets D008 (5'- ATAATGCTAAATTAGACAACTTGAGCGA-3') and R008 (5' TTAGAATAATCAACAAACATCACGCCAT-3')¹³. The amplification product of cagA gene was 297 base pairs (bp) in length.

The PCR reaction mixtures were prepared by using TopTaq Master Mix Kit (Qiagen, Germany) in a final volume of 25 μ l containing 1.25 units TopTaq DNA polymerase, 1 X PCR buffer, 1.5 mM MgCl2 and 200 μ M of each dNTP, 0.2 μ M of each primer, 10 μ l of molecular grade water and 2.5 μ l of DNA. The mixtures were placed in a PCR thermocycler (Eppendorf, Germany). The PCR conditions included an initial denaturation of target DNA at 94°C for 1 min, followed by 35 cycles of denaturation at 94°C for 1 min, primer annealing at 58°C for 1 min, extension at 72°C for 1 min and final extension at 72°C for 15 min.

PCR products were run on 1.5% agarose gels in TBE buffer according to the manufacturer's instructions and it revealed presence of *H. pylori* cagA virulence gene indicating 297 bp (Figure 1).

DISCUSSION

The incidence of antibiotic resistance differs from one geographic area to another. It is high in developing countries compared to developed countries^[4]. In Malaysia Metronidazole is used as first-line therapy for *H. pylori* infection with highest resistance of (40%) reported^[5]. High prevalence of Metronidazole resistance has also been reported in India (90%)^[6].

The relationship between the success or failure of *H. pylori* eradication therapy and cagA status has been explained by the enhanced gastric mucosal inflammation. Patients having severe inflammatory cell infiltrations in the antral mucosa were associated with higher cure rates¹⁷⁾, therefore the presence of cagA might not be a risk factor in development of Metronidazole resistance¹⁸⁾. In contrast, our case which is cagA positive is resistant to Metronidazole therapy. The continued resistance observed in this patient might be due to combination of bacterial, environmental and genetic factors. Further studies need to be done especially in areas where the resistance rate is high.

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REFERENCES

- Suzuki RB, Cola RF, Cola LT, Ferrari CG, Ellinger F, Therezo AL, et al. Different risk factors influence peptic ulcer disease development in a Brazilian population. World J Gastroenterol 2012; 18: 5404-5411.
- Amieva MR, El-Omar EM. Host-bacterial interactions in Helicobacter pylori infection. Gastroenterology 2008; 134:306-323.
- 3) Ibrahim IAA., Kamisah Y, Nafeeza MI, Nur Azlina MF. Modulation of Gastric Motility and Gastric Lesion Formation in Stressed Rats Given Enteral Supplementation of Palm Vitamin E and a - Tocopherol. IMJ 2011; 18: 47-52.
- Megraud F, Lehours P. Helicobacter pylori detection and antimicrobial susceptibility testing. Clin Microbiol Rev 2007; 20: 280-322.
- Rahman SH, Azam MG, Rahman MA, Arfin MS, Alam MM, Bhuiyan TM, et al. Noninvasive diagnosis of H pylori infection: evaluation of serological tests with and without current infection marker CIM. World J Gastroenterol 2008; 14: 1231-1236.
- 6) Kopanski Z, Wasilewska-Radwanska M, Jung A, Kuc T, Schlegel-Zawadzka M, Witkowska B. et al. Diagnostic value of the urine test with 14C-urea in the detection of the Helicobacter pylori infection. IMJ 1999; 6: 109-112.
- Rautelin H, Lehours P, Megraud F. Diagnosis of Helicobacter pylori infection. Helicobacter 2003; 8 (Suppl 1): 13-20.
- Malfertheiner P, Megraud F, O'Morain C, Bazzoli F, El-Omar E, Graham D, et al. Current concepts in the management of Helicobacter pylori infection: the Maastricht III Consensus Report. Gut 2007; 56: 772-781.
- Tan HJ, Rizal AM, Rosmadi MY, Goh KL. Distribution of Helicobacter pylori cagA, cagE and vacA in different ethnic groups in Kuala Lumpur, Malaysia. J Gastroenterol Hepatol 2005; 20: 589-594.
- 10) Ramelah M, Aminuddin A, Alfizah H, Isa MR, Jasmi AY, Tan HJ, et al. cagA gene variants in Malaysian Helicobacter pylori strains isolated from patients of different ethnic groups. FEMS Immunol Med Microbiol 2005; 44: 239-242.
- Kuipers EJ, Perez-Perez GI, Meuwissen SG, Blaser MJ. Helicobacter pylori and atrophic gastritis: importance of the cagA status. J Natl Cancer Inst 1995; 87: 1777-1780.
- 12) Kao CY, Sheu BS, Sheu SM, Yang HB, Chang WL, Cheng HC, et al. Higher motility enhances bacterial density and inflammatory response in dyspeptic patients infected with Helicobacter pylori. Helicobacter 2012; 17: 411-416.
- 13) Sillakivi T, Aro H, Ustav M, Peetsalu M, Peetsalu A, Mikelsaar M. Diversity of Helicobacter pylori genotypes among Estonian and Russian patients with perforated peptic ulcer, living in Southern Estonia. FEMS Microbiol Lett 2001; 195: 29-33.
- Megraud F. H pylori antibiotic resistance: prevalence, importance, and advances in testing. Gut 2004; 53: 1374-1384.
- 15) Salasawati H, Ramelah M, Jasmi A, Mazlan N, Isa R, Alfizah H, et al. Antibiotic resistance of Helicobacter pylori: association with gastroduodenal disease in Malaysia. Med J Malaysia 2001; 56(Suppl A): 65.
- Chowdhury A, Berg DE, Jeong JY, Mukhopadhyay AK, Nair GB. Metronidazole resistance in *Helicobacter pylori*: magnitude, mechanism and implications for India Indian J Gastroenterol 2002: 21: 23-28.
- 17) Zanten SJ, Bradette M, Farley A, Leddin D, Lind T, Unge P, et al. The DU-MACH study: eradication of Helicobacter pylori and ulcer healing in patients with acute duodenal ulcer using omeprazole based triple therapy. Aliment Pharmacol Ther 1999; 13: 289-295.
- 18) Taneike I, Nami A, O'Connor A, Fitzgerald N, Murphy P, Qasim A, et al. Analysis of drug resistance and virulence-factor genotype of Irish Helicobacter pylori strains: is there any relationship between resistance to metronidazole and cagA status? Aliment Pharmacol Ther 2009; 30: 784-790.